

modification and rescue might involve multiple *Wolbachia* genes.

The existence of *Wolbachia* strains that do not modify sperm but can rescue the cytoplasmic incompatibility phenotype has been predicted by theory<sup>13,14</sup>. Our work suggests that previously classified  $\text{mod}^- \text{resc}^-$  strains of *Wolbachia* can in fact rescue cytoplasmic incompatibility and can therefore be considered  $\text{mod}^- \text{resc}^+$ . This finding helps explain how *Wolbachia*  $\text{mod}^-$  strains could have spread into insect populations without the action of cytoplasmic incompatibility. *Wolbachia* strains that can rescue but not cause cytoplasmic incompatibility can be expected to increase in frequency in populations by essentially parasitizing the cytoplasmic incompatibility sperm modification effect of a related *Wolbachia* strain.

The potentially lower fitness costs and perfect maternal transmission of these strains<sup>7,9</sup> might explain how they could displace the initial strain that they parasitized in order to invade the host population. This is in accordance with models predicting that *Wolbachia* will evolve towards neutrality with respect to cytoplasmic incompatibility as long as maternal transmission frequency can be increased<sup>13</sup>.

**Kostas Bourtzis\***, **Stephen L. Dobson**,  
**Henk R. Braig**, **Scott L. O'Neill**

Section of Vector Biology,  
Department of Epidemiology and Public Health,  
Yale University School of Medicine,  
60 College Street, New Haven, CT 06510, USA  
e-mail: scott.oneill@yale.edu

\*Alternative address: Institute of Molecular Biology  
& Biotechnology, Foundation for Research and  
Technology — Hellas, Heraklion, Crete, Greece

## ...and discovered on Mount Kilimanjaro

The endocyttoplasmic bacterium *Wolbachia* causes the death of arthropod embryos, when present in reproductive organs, by cytoplasmic incompatibility<sup>1</sup>. *Wolbachia* is harboured in both sexes but transmission is maternal only. Cytoplasmic incompatibility occurs when infected males inseminate uninfected females<sup>2</sup> or females bearing a different variant of *Wolbachia*<sup>3,4</sup>. Two kinds of *Wolbachia* have been described<sup>1</sup>: ( $\text{mod}^+ \text{resc}^+$ ) which induces cytoplasmic incompatibility by modifying sperm but can rescue this phenotype when in the egg, and ( $\text{mod}^- \text{resc}^-$ ) which neither induces cytoplasmic incompatibility nor rescues from it. Theory predicts a third kind ( $\text{mod}^- \text{resc}^+$ ) that would not induce cytoplasmic incompatibility but would rescue from it<sup>5-7</sup>. We have found such a *Wolbachia* variant in a *Drosophila simulans* population on Mt Kilimanjaro, Tanzania. The existence of a  $\text{mod}^- \text{resc}^+$  *Wolbachia* shows that the modification and rescue functions can be dissociated with regard to the cytoplasmic incompatibility process.

We detected this *Wolbachia* variant ( $wKi$ ) in isofemale lines from flies captured in March 1996. Of 49 lines, 9 showed a positive PCR signal with primers for the *Wolbachia* 16S rRNA gene<sup>8</sup>. Infected ( $Kili^+$ ) and uninfected ( $Kili^-$ ) Kilimanjaro males were crossed with  $Kili^-$  females. No cytoplasmic incompatibility was detected. The percentage of unhatched eggs did not differ significantly whether the males were infected or uninfected ( $12.3 \pm 2.6\%$  and  $10.7 \pm 3.3\%$ , respectively).

We tested the compatibility of  $Kili^+$  flies with individuals infected by each of the three natural  $\text{mod}^+ \text{resc}^+$  *Wolbachia* strains ( $wRi$ ,  $wHa$  or  $wNo$ )<sup>9</sup>.  $Kili^+$  males were compatible with females infected by  $wRi$  ( $12.0 \pm 5.1\%$  unhatched eggs),  $wHa$  ( $14.0 \pm 4.1\%$ ) or  $wNo$  ( $8.5 \pm 1.3\%$ ).  $Kili^+$  females were incompatible with males infected by  $wRi$  (100.0% unhatched eggs) or  $wHa$  ( $84.7 \pm 3.3\%$ ). Surprisingly, the  $Kili^+$  females were completely compatible with  $wNo$ -infected males ( $8.5 \pm 2.1\%$  unhatched eggs), whereas  $Kili^-$  females were incompatible ( $70.2 \pm 3.5\%$ ).  $wKi$  thus behaves as a  $\text{mod}^- \text{resc}^+$  variant with regard to the  $wNo$  type.

Cytoplasmic incompatibility helps *Wolbachia* in invading previously uninfected populations, because it eliminates uninfected eggs, giving a higher fitness to infected females. However, during the invasion process, any  $\text{mod}^-$  variant arising by mutation will also increase in frequency as long as it is still  $\text{resc}^+$ . Under certain conditions,

these  $\text{mod}^- \text{resc}^+$  variants would completely replace the  $\text{mod}^+ \text{resc}^+$  wild type. If this happened, the population would again become vulnerable to invasion by uninfected cytoplasm, because  $\text{mod}^-$  mutants do not exert any selective pressure against uninfected eggs. The infection might then be lost with time. Meanwhile, because  $\text{mod}^+$  variants have been eliminated from the population, the rescue capability becomes useless and  $\text{resc}^-$  mutants can be expected to arise.

The *Wolbachia* variants discovered in wild *D. simulans* populations<sup>9,10</sup> have been of two types:  $\text{mod}^+ \text{resc}^+$  ( $wRi$ ,  $wHa$ ,  $wNo$ ) and  $\text{mod}^- \text{resc}^-$  ( $wMa$ ,  $wA$ ).  $wA$  has been found both on the American and Australian continents, whereas  $wMa$  is known only in Madagascar. We established that  $wA$  and  $wKi$  are different because  $wA$  cannot rescue from the  $wNo$  cytoplasmic incompatibility phenotype<sup>10</sup>. The status of  $wMa$  is unknown in this respect, so  $wMa$  and  $wKi$  might be the same variant. Moreover, the 16S rDNA amplification product of  $wKi$  has one *VspI* restriction site, as do  $wMa$  and  $wNo$  sequences<sup>8</sup>, but such a site is absent from  $wRi$ ,  $wHa$  and  $wA$  sequences.

Eastern Africa is thought to be the centre of origin of *D. simulans*<sup>11</sup>, so the *Wolbachia* infection of the Kilimanjaro population could be one of the oldest in this species, implying that  $\text{mod}^- \text{resc}^+$  variants derive from a  $\text{mod}^+ \text{resc}^+$  ancestor with time.

In contrast,  $wNo$  is known only in the Seychelles and in New Caledonia<sup>10</sup>, so perhaps  $wNo$ -infected populations originate from migrants that left East Africa bearing the  $\text{mod}^+ \text{resc}^+$  ancestor. After geographic isolation, the Indo-Pacific variant could have remained  $\text{mod}^+ \text{resc}^+$  while the ancestral variant has evolved into  $\text{mod}^- \text{resc}^+$  in Africa. The maintenance of the rescue function implies that it might have a further role.

The proportion of insect species potentially infected by *Wolbachia* is estimated at 16% (ref. 1), so further research might yield other  $\text{mod}^- \text{resc}^+$  variants. Mt Kilimanjaro is probably just the tip of an iceberg.

**Hervé Mercot**, **Denis Poinso**

Laboratoire Dynamique du Génome et Évolution,  
Institut Jacques Monod, CNRS-Université Paris 7,  
75251 Paris, France  
e-mail: mercot@ccr.jussieu.fr

1. Werren, J. H., Windsor, D. & Guo, L. R. *Proc. R. Soc. Lond. B* **262**, 197–204 (1995).
2. O'Neill, S. L., Hoffmann, A. A. & Werren, J. H. *Infectious Passengers: Inherited Microorganisms and Arthropod Reproduction* (Oxford Univ. Press, Oxford, 1997).
3. Callaini, G., Dallai, R. & Riparbelli, M. G. *J. Cell Sci.* **110**, 271–280 (1997).
4. Callaini, G., Riparbelli, M. G., Giordano, R. & Dallai, R. *J. Invert. Pathol.* **67**, 55–64 (1996).
5. O'Neill, S. L. & Karr, T. L. *Nature* **348**, 178–180 (1990).
6. Hoffmann, A. A., Turelli, M. & Simmons, G. M. *Evolution* **40**, 692–701 (1986).
7. Hoffmann, A. A., Clancy, D. & Duncan, J. *Heredity* **76**, 1–8 (1996).
8. Holden, P. R., Jones, P. & Brookfield, J. F. *Genet. Res.* **62**, 23–29 (1993).
9. Giordano, R., O'Neill, S. L. & Robertson, H. M. *Genetics* **140**, 1307–1317 (1995).
10. Montchamp-Moreau, C., Ferveur, J. & Jacques, M. *Genetics* **129**, 399–407 (1991).
11. Zhou, W., Rousset, F. & O'Neill, S. L. *Proc. R. Soc. London B* (in the press).
12. Bourtzis, K., Nirgianaki, A., Onyango, P. & Savakis, C. *Insect Mol. Biol.* **3**, 131–142 (1994).
13. Turelli, M. *Evolution* **48**, 1500–1513 (1994).
14. Werren, J. H. & O'Neill, S. L. in *Infectious Passengers: Inherited Microorganisms and Arthropod Reproduction* (eds O'Neill, S. L., Hoffmann, A. A. & Werren, J. H.) 1–41 (Oxford Univ. Press, Oxford, 1997).
15. Bourtzis, K., Nirgianaki, A., Markakis, G. & Savakis, C. *Genetics* **144**, 1063–1073 (1996).
16. Mercot, H., Llorente, B., Jacques, M., Atlan, A. & Montchamp-Moreau, C. *Genetics* **141**, 1015–1023 (1995).

1. Werren, J. H. *Annu. Rev. Entomol.* **42**, 587–609 (1997).
2. Hoffmann, A. A., Turelli, M. & Simmons, G. M. *Evolution* **40**, 692–701.
3. Breeuwer, J. A. J. & Werren, J. H. *Nature* **346**, 558–560 (1990).
4. O'Neill, S. L. & Karr, T. L. *Nature* **348**, 178–180 (1990).
5. Prout, T. *Evolution* **48**, 909–911 (1994).
6. Turelli, M. *Evolution* **48**, 1500–1513 (1994).
7. Hurst, L. D. & McVein, G. T. *Proc. R. Soc. Lond. B* **263**, 97–104 (1996).
8. Rousset, F., Vautrin, D. & Solignac, M. *Proc. R. Soc. Lond. B* **247**, 163–168 (1992).
9. Clancy, D. J. & Hoffmann, A. A. *Trends Ecol. Evol.* **11**, 145–146 (1996).
10. Mercot, H. & Poinso, D. *Entomol. Exp. et Appl.* (in the press).
11. Lachaise, D. *et al. Evol. Biol.* **22**, 159–225 (1988).